in chloroform. NMR solutions of $Zn(II)$ [14]12eneN₄(amine) in chloroform- d_1 slowly decompose (hours) to give the dideuterated form of the ligand, $\overline{D_2}$ [14] 12eneN₄, and a white solid (possibly $ZnCl₂$).

Both the manganese and zinc complexes are sensitive to alkylating agents. Addition of excess $CH₃X$, where X is Iand FSO₃⁻, to benzene solutions containing the complexes leads to their destruction. However, the only alkylated product which could be identified from the reaction mixture was [CH3NR3]X. Attempts to isolate the alkylated form of the ligand lead to hygroscopic yellow solids which could not be characterized. The compounds are easily decomposed by aqueous strong mineral acids, e.g., HCl and HC104. Unlike the Mn(II1) derivatives addition of Et3N will not restore the original complex. However, if anhydrous $HSO₃CF₃$ is added to acetonitrile suspensions of the compounds under nitrogen a dissolution accompanied by a color change occurs. Addition of triethylamine results in the regeneration of $M(II)[14]$ -12eneN4(amine) (eq 2).

The synthesis and properties of manganese complexes containing other unsaturated tetraaza and mixed N-0 donor ligands will be subsequently reported.

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Registry No. Hz[14] 12eneN4, 56276-51-8; Zn(II)[14]- 12ene $N_4(N(Et)3)$, 56943-15-8; Zn(II)[14]12ene $N_4(N(n-Pr)3)$, 56943-16-9; Mn(II)[14]12eneN₄(N(Et)3), 56943-17-0; Mn(II)- $[14]12$ eneN₄(N(n-Pr)3), 56943-18-1; Mn(III)[14]12eneN₄(Cl), 56943-19-2; Mn(III)[14] 12eneN4(Br), 56943-20-5; Mn(III)[14]- 12eneN4(NCS), 56943-21-6; Mn(III)[141 12eneN4(N3), 56943-23-8; Mn(III)(acac)zCI, 51 197-21-8; Mn(III)(acac)zBr, 51 147-59-2; Mn(III)(acac)₂(NCS), 52242-31-6; Mn(III)(acac)₂(N₃), 52242-29-2.

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Carbon- 13 Nuclear Magnetic Resonance of Substituted Tetraphenyl Porphyrins and Their Complexes with Ruthenium, Indium, and Titanium

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¹³C NMR spectra of H₂P and MPXY (P = tetrakis(p-trifluoromethylphenyl)porphyrin, tetrakis(p-isopropylphenyl)porphyrin, and **tetrakis(o-tolyl)porphyrin, M = Ru, X = CO, Y = tetrahydrofuran, 4-tert-butylpyridine, or CO; M = In, X = Cl;** $M = Ti$, $X = O$; $P = octaethylporphyrin$, $M = Ru$, $X = CO$, $Y = tetrahydrofuran$; $M = In$, $X = Cl$) and $RuPX₂$ ($P = O$ $tetrakis(p-trifluorometry|phenyl)porphyrin)$, $X = trimethylt phosphite and *tert*-butyl isocyanide) are reported and interpreted.$ Chemical shift differences for nonequivalent sites and ring current effects are compared for IH and 13C NMR spectra. Significant dependence of porphyrin chemical shifts on the metal is observed.

Introduction

Recently, increasing effort has been applied to understanding and using ¹³C NMR to elucidate the chemistry of porphyrins and metalloporphyrins. These studies have concerned the apparent electron delocalization pathway in natural porphyrins,¹ biosynthesis,² identification of type isomers,³ isotropic shifts in paramagnetic iron(II1) porphyrins,4 ring current effects on the porphyrin resonances and on the resonances of other coordinated ligands,5-6 assignment of spectra of tetrapyrroles,⁷ and Cd(II), $Hg(II)$, and Tl(III) complexes of tetraphenylporphyrin.3c We now report the I3C NMR spectra of $Ru(II)$, $(TiO)^{2+}$, and $In(III)$ complexes of sub-

stituted tetraphenylporphyrins. The compounds chosen for study provide comparison of the relative IH and 13C chemical shift differences for sites which are nonequivalent due to restricted phenyl ring rotation.⁸ These compounds also provide an indication of the effect of complexed metal on porphyrin chemical shifts.

Experimental Section

Physical Measurements. Infrared spectra were recorded as Nujol or halocarbon mulls on a Perkin-Elmer 710 grating spectrometer. Visible spectra were obtained in chloroform solution on a Beckman Acta V spectrometer. Data are given below with wavelengths in nanometers and $log \epsilon$ in parentheses. ¹H NMR spectra were run at

power levels well below saturation on a Varian HA-100 spectrometer with the spectrometer locked on the solvent resonance. Unless otherwise noted, IH chemical shifts are reported in parts per million downfield of tetramethylsilane (TMS) for 1,1,2,2-tetrachloroethane solutions at ambient temperature. ¹³C NMR spectra were obtained on a Varian CFT-20 13C pulsed Fourier transform NMR spectrometer operating at 20 MHz with proton noise decoupling and internal deuterium lock. Typically, 4096 data points were taken over a 4-kHz spectral width using a ca. 20° pulse and a ca. 1 sec acquisition time. Samples containing 40-100 mg of compound in 1.5 ml of CDCl₃ in 10-mm tubes were run at a probe temperature of ca. 35°. ¹³C chemical shifts are reported relative to TMS, using the central line of the CDCl₃ triplet as a secondary standard with a shift of 77.05 ppm. Positive shifts indicate lower shielding. Shifts are considered accurate to ± 0.1 PPm.

The spectra of the ruthenium carbonyl porphyrins were first run on the compounds as isolated. A second spectrum was obtained in the presence of excess 90% enriched 13CO (Merck). CO exchange affected ¹³C enrichment of the coordinated CO allowing facile detection of the CO resonance which was difficult to observe in natural abundance.

Preparation **of** Compounds. **Ru(p-CF3-TPP)(P(OMe)3)2.9** Addition of excess trimethyl phosphite (P(OMe)3) to 100 mg of Ru- $(CO)(p-CF_3-TPP)(THF)^{8b}$ dissolved in 2 ml of trichloroethylene caused an immediate color change from red to red-brown. The solution was stirred for 2 hr. Reduction of volume and addition of heptane afforded blue-purple crystals. Conversion was quantitative: the ir showed no CO stretch; ¹H NMR, +8.28, (pyrrole-H, singlet), +8.17 $(o-H, doublet, J = 7.5 Hz), +7.91$ (*m*-H, doublet, $J = 7.5 Hz$), $+0.85$ (CH₃, apparent triplet, $J = 5$ Hz); visible spectrum 405 sh (4.71), 426 (5.55), 527 (4.09). Anal. Calcd for C54H42N4F12RuO6P2: C, 52.56; H, 3.43; N, 4.54; P, 5.25. Found: C, 52.75; H, 3.22; N, 4.80; P, 5.25.

Ru(p-CF3-TPP)(t-Bu-NC)za Addition of 0. I ml of t-Bu-NClo to 100 mg of $Ru(CO)(p-CF_3-TPP)(THF)^{8b}$ dissolved in 2 ml of trichloroethylene caused vigorous bubbling and the solution turned darker. After stirring for 1 hr, the volume was reduced and heptane was added. Blue-purple crystals formed on standing. Conversion was quantitative: ir ν CN 2140 cm⁻¹, no CO band; ¹H NMR +8.34 (pyrrole-H, singlet), +8.25 (o-H, doublet, $J = 8$ Hz), +7.94, (*m*-H, doublet, $J = 8$ Hz), -0.48 (CH₃, singlet); visible spectrum 404 sh $(4.66), 421 (5.67), 525 (4.09).$ Anal. Calcd for C58H42N6F12Ru: C, 60.46; H, 3.68; N, 7.30. Found: C, 59.92; H, 3.82; **N,** 7.36.

Synthesis and characterization of the remaining metalloporphyrins are reported elsewhere.8

Assignment **of** the I3C NMR Resonances. The numbering of the carbon atoms for all of the tetraphenylporphyrin derivatives is indicated in I. The chemical shifts are given in Table I. In discussing

assignments, average values for a porphyrin and its complexes are used; see Table **1.**

 $H_2(p$ -CF₃-TPP) and Complexes. The ¹³C chemical shifts and C-F coupling constants reported for α, α, α -trifluoromethyltoluene^{11a} were used to assign the resonances in the ¹³C spectra of $H_2(p$ -CF₃-TPP) and its complexes. The J_{C-F} for CF_3 , C_4 , and C_3 in the protondecoupled spectra allow unambiguous assignment of the resonances for these carbons. In $H_2(p-CF_3-TPP)$ one of the central lines of the quartets assigned to C_4 and CF_3 is obscured by β -C so the centers of the quartets were estimated using the coupling constants observed

t previous assignments for M(TPP), $M = Cd(II),3c$ Hg(II), $3c$ Tl(III), $3c$ for the complexes. The quaternary and nonquaternary carbons are readily distinguished on the basis of intensity differences under the experimental conditions used. The only nonquaternary carbons not assigned on the basis of C-F coupling constants are β -C and C₂. The signal at 134.5 ppm is assigned to C_2 since it splits into two signals of equal intensity in the complexes. This splitting is expected since C2 and C2' are nonequivalent in these complexes due to restricted rotation of phenyl rings and slow axial ligand exchange.8 The remaining nonquaternary signal, at 132.1 ppm, is therefore assigned to β -C. Of the quaternary carbon signals, the one due to α -C is easily assigned since it broadens in the free porphyrin due to N-H tautomerism.^{3b,12} Two quaternary carbons, C_1 and meso-C, remain to be assigned to the signals at 120.7 and 146.0. Based on the chemical shift of the substituted phenyl carbon in biphenyl (141.7 ppm) , 13a styrene (138.2 ppm),l3band 3-phenylpyrrole **(135.5** ppm)l4 and the 3-C in 3-phenylpyrrole (123.8 ppm) , ¹⁴ it seems reasonable to assign the signal at 146.0 ppm to C_1 and the signal at 120.7 ppm to meso-C. These assignments are considered further in the comparison with the and $Zn(II)$, ^{4b}

 $H_2(p-i-Pr-TPP)$ and Complexes. Assuming the usual additivity of aromatic ring substituent effects, the phenyl ring resonances of $H_2(p-i-Pr-TPP)$ and its complexes can be assigned by comparison with the spectra of substituted benzenes in Table I1 and the spectra of $H₂(p-CF₃-TPP)$ and its complexes. The chemical shift differences in ppm between isopropylbenzene and trifluoromethylbenzene (Table **11)** are Ca, 17.9; Cb, 1.1; Cc, -0.4; Cd, -5.8. The proposed assignments in Table I result in the following differences in average chemical shifts between $H_2(p-i-Pr-TPP)$ and its complexes and $H_2(p-CF_3-TPP)$ and its complexes $(C_4(C_a), 18.1; C_3(C_b), 1.0; C_2(C_c), 0.1; C_1(C_d), -6.5)$, closely paralleling the values for the substituted benzenes. The α -C is readily assigned on the basis of the broadening in the spectrum of the free porphyrin. The only nonquaternary carbon remaining to be assigned, β -C, is assigned on the basis of intensity to the resonance at 132.1 ppm. The remaining signal at 121.9 ppm is meso-C.

 $H₂(o-Me-TPP)$ and Complexes. The additivity of aromatic ring substituent effects on 13C NMR chemical shifts permits calculation of the shifts expected for $H_2(o-Me-TPP)$ using the data in Table II, the assignments made above for $H_2(p-CF_3-TPP)$ and $H_2(p-i-Pr-TPP)$ and their complexes, and the substituent effect of a methyl group.16 First, the shifts for $H_2(p-CF_3-TPP)$ and $H_2(p-i-Pr-TPP)$ respectively yield the following predictions for H₂(TPP): (a) C₁, 142.8; C₂, 134.3; C3, 127.0; C4, 127.8; and (b) Ci, 142.1; C2, 134.8; C3, 126.9; C4, 128.0 ppm. The average of these two sets of predicted shifts agrees closely with the values reported for $M(TPP)^{3c,4b}$ except for C₁ in Zn(TPP).^{4b} If the methyl group is now put **on** C2', the substituent effect of a methyl group¹⁶ predicts C₁, 143.1; C₂, 134.5; C₂', 143.5; C₃, 124.1; C₃', 127.7; C₄, 127.8. The observed spectra can be assigned with C_1 , 141.3; C₂, 134.0; C2', 139.9; C3, 124.4; C3', 129.4; C4, 128.7 ppm, which is in good agreement with the predictions, particularly if the small upfield shift in C_1 and C_2 ' is attributed to a strain contribution. The possible ambiguity between C_3 ' and C_4 is resolved by the nonequivalence observed in C₃' due to restricted rotation of the phenyl rings.⁸ α -C, β -C, and meso-C are assigned by arguments similar to those used for $H_2(p-i-Pr-TPP)$ and complexes.

OEP Complexes. The resonances of the octaethylporphyrin complexes were assigned by analogy with the reported spectra of $H₂(OEP)$ and $Zn(OEP)$.^{3b}

Discussion

Comparison with Previous Assignments. The assignments given for C_1 and meso-C are reversed from those chosen previously for H2(TPP) and Zn(TPP).4b Our assignments are based on the following grounds. (1) The assigned chemical shifts of C₁ are consistent with chemical shifts in biphenyl,^{13a} styrene,^{13b} and 3-phenylpyrrole.¹⁴ (2) The variation of the signals at 139-147 ppm, assigned to C_1 , with changes in phenyl ring substituent is consistent with additivity rules for aromatic rings whereas the signal at 120.7-121,9 ppm, assigned to meso-C, is much less dependent on phenyl ring substituent. (3) The difference in chemical shift between the 3-carbon in pyrrole'3c and in 3-phenylpyrrole14 is +16.1 ppm. The chemical shift difference between benzenel3b and the substituted carbon in biphenyl^{13a} is 13.3 ppm. Thus, replacement of a proton on an aromatic ring by a phenyl ring appears to

 a_{Estimate} by comparison with complexes. B-C obscures part of each quartet in free porphyrin. DNot observed. CSpectrum in d_a-pyridine/D₂0 from reference 4b. ^dSpectrum in CDC1₃ with ca. 2 equiv. of pyrrolidi**ne a**dded, from reference 3c. ^eSpectrum in CDC1₃ from reference 3b. ^fNot reported. ⁹Assignments may be interchanged.

Table II. ¹³C Chemical Shifts of Substituted Benzenes

deshield the substituted carbon by ca. 13-16 ppm. The chemical shift difference of 23-24 ppm between resonances for meso-C in OEP and tetraphenylporphyrin complexes (after allowing for ring substituent effects) would seem more consistent with this expectation than the previously proposed 45-ppm difference.^{4b} The assignments are consistent with those reported for $M(TPP)$, $M = Cd(II)$, $Hg(II)$, and Tl(III), on the basis of single-frequency off-resonance decoupling^{3c} after completion of this work.

The proposed chemical shift assignments in $H_2(o$ -Me-TPP) and complexes for carbons C₂ and C₃ agree with those proposed by Abraham et al. for $H_2(o$ -Me-TPP) and its Tl(III) complex.^{3c} The assignments for C_3 and C_4 are reversed in this paper relative to those of Abraham et al.^{3c} as are the assignments for C_1 and C_2 ^t. In each case the two resonances concerned are within 1 ppm of each other. However, the resonances assigned in this work to C_2 ['] and C_3 ['] in $H_2(o-$ Me-TPP) split into pairs of signals in the metal complexes as expected for nonequivalent ortho and meta carbons while the signals for C_1 and C_4 remain unsplit. Thus comparison of the spectrum of the free porphyrin with spectra of the metal complexes removes the possible ambiguity.

Dependence of Chemical Shifts on Coordinated Metal. In NMR studies of paramagnetic metal complexes, calculation of the isotropic shift requires an assumed chemical shift for the same atom in the absence of paramagnetism. Zinc complexes of the same ligand are commonly used as diamagnetic reference compounds. The uncertainty introduced by this assumption can only be estimated by considering the range of shifts for the same ligand atom in complexes with various diamagnetic metals.

The series of compounds reported herein yields information pertinent to estimating the uncertainties in isotropic shift calculations for paramagnetic metalloporphyrins. The ¹³C chemical shift of the α -C is particularly sensitive to changes in metal ion. For a given substituted tetraphenylporphyrin we observe that different metals cause a range of ca. 6 ppm in the ¹³C chemical shift of α -C, 1.5 ppm for β -C, 2 ppm for meso-C, 2 ppm for C_1 , 0.5 ppm for C_2 or C_3 , and 1.5 ppm for C₄. In the ¹H NMR spectra of the same complexes the range of shifts are 1 ppm for pyrrole-H, 0.3 ppm for o-H, and 0.2 ppm for m-H. Similarly, with OEP complexes, different metals cause a range of ca. *5* ppm in the 13C chemical shift of the α -C, 2 ppm for β -C, 2 ppm for meso-C, 0.2 ppm for $CH₂$, and 0.3 ppm for CH₃. In the ¹H NMR of the same complexes the range of shifts are 0.6 ppm for meso-H, 0.3 ppm for CH2, and 0.3 ppm for CH3.

In some cases the range of chemical shifts observed for the few metals studied is as large as the isotropic shifts reported for paramagnetic metalloporphyrins. For example, the isotropic shifts of ¹³C and ¹H resonances in Fe(TPP)(CN)₂ reported4b relative to Zn(TPP) as diamagnetic reference are listed below, together with the uncertainties calculated using the above ranges of chemical shifts for diamagnetic metalloporphyrins: pyrrole-H, 19.1 (5%); m-H, 0.2 (100%); o-H, C₂, -33.3 (2%); α -C, 111.0 (5%); meso-C, 24.5 (8%); and C₁, -11.2 (18%). A similar analysis of the isotropic shifts for $Fe(TPP)Cl$ and $Fe(OEP)Cl$,¹⁷ referenced against the diamagnetic nickel porphyrin, indicates possible uncertainties of up to 10% from this source. Clearly the choice of diamagnetic reference can strongly affect the results of isotropic shift calculations and must be very carefully justified for any particular system. Both the relative range of shifts and the resultant uncertainty in the isotropic shifts are of the same order of magnitude for 13C and 1H NMR spectra. 3.0 (10%); P-C, 41.9 *(5%);* C4, -1.0 (67%); C3, -0.5 (100%);

Carbonyl Resonances. The carbonyl resonance was difficult to detect in the natural abundance spectra. Addition of excess 13CO to the sample tube resulted in substantial isotopic enrichment and the CO resonance became the largest peak in the spectrum. All other resonances in the spectrum of Ru- $(CO)(p-i-Pr-TPP)(py)$ were unchanged by the addition of ¹³CO. In the spectrum of $Ru(CO)(p-CF_3-TPP)(THF)$ separate resonances are observed for the nonequivalent C₂ and C_2 ' and a broadened multiplet was observed for C_3 and C_3 ' due to restricted rotation of phenyl rings and slow axial ligand exchange.⁸ In the presence of excess $13CO$ the C₂, C₂' resonances collapse to a single resonance and C_3 , C_3 ' collapse to a quartet (due to coupling to CF3) indicative of a fast CO exchange from one side of the porphyrin plane to the other. The same spectral changes would be expected for the permutationally equivalent process of phenyl ring rotation, but ring rotation has been demonstrated to be slow on the NMR time scale at room temperature for a range of ruthenium porphyrin complexes.8 All other resonances are unchanged by CO addition. If the excess CO is then allowed to escape from the sample tube, the CO resonance moves 0.5 ppm to lower shielding relative to the shift observed in the presence of excess CO (179.6 to 180.1 pprn). The remainder of the spectrum is the same as in the presence of excess CO. Apparently CO can compete, on the NMR time scale, with THF for a coordination site in $Ru(CO)(p-CF_3-TPP)(THF)$ but cannot compete, on the NMR time scale at 35°, with pyridine for a coordination site in $Ru(CO)(p-i-Pr-TPP)(py)$. Upon addition of CO to a sample of Ru(CO)(OEP)(THF), the THF resonances are shifted to lower shielding $(1.3$ ppm for $C_{2.5}$ and 3.0 ppm for $C_{3,4}$). From the ¹H NMR spectra it is known that exchange between free and coordinated THF is fast on the NMR time scale at room temperature. The shift in the THF resonances in the presence of excess CO is interpreted as indicating partial displacement of coordinated THF due to dicarbonyl formation. The CO resonance in the presence of excess CO is 2 ppm to greater shielding of the resonance in the absence of excess CO. All other resonances in the spectrum of Ru(CO)(OEP)(THF) are unchanged by the addition of CO.

Separate signals are not observed for mono- and dicarbonyl species, or for uncoordinated ^{13}CO in solution. Apparently, CO exchange is rapid under the conditions of the measurement and the observed shift is a weighted average for the various CO species present. Under the conditions of the spectra, the concentration of free CO in solution was, at most, ca. 3-7% of the concentration of porphyrin.18 The reported chemical shifts for CO are 182.2 ppm (gas phase)^{19a} and 184.6 ppm $(D_2O$ solution).^{19b} The ¹³CO shifts in the Ru(CO)(porphyrin) complexes are in the range 179.6-183 ppm, very close to the shift anticipated for free CO. Thus the relative amount of excess CO in solution would not be expected to have a major effect on the observed average chemical shift. With the caution of these uncertainties it is interesting to note that the larger difference in CO chemical shift in the presence and absence of CO for $Ru(CO)(OEP)(L)$ than for $Ru(CO)(p-CF)$ -TPP)(L) is consistent with considerably greater dicarbonyl formation for the former than for the latter. The difference in behavior of the two ruthenium porphyrins is consistent with the previous findings that the second carbonyl is bound more strongly in $Ru(CO)_{2}(OEP)$ than in $Ru(CO)_{2}(p\text{-}CF_{3}\text{-}TPP).^{20}$ stry, *Vol. 15*, *No. 1*, 1976 137
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Previously reported 13C shifts for carbonyl groups bound to ruthenium range from +237.0 ppm for the average CO environment in $(C_5H_5)(CO)Ru(\mu$ -CO $)$ ₂Ru(CO)(C₅H₅)²¹ to +189.3 and 190.1 ppm in $Ru_3(CO)_9(CMe)H_3$.²² Other values include +199.7 ppm for the average environment in Ru₃(CO)₁₂,²³ 192.1-199.7 ppm in Ru₃(CO)⁹(C₆H₉)H₂₄ and 193.7-198.3 ppm in cis-RuCl₂(CO)₂(L)₂ where L is a variety of alkyl phosphines.25 The 13CO resonances in the ruthenium porphyrin complexes appear to be more shielded than those previously reported. The diamagnetic anisotropy of the porphyrin ring may contribute substantially to this difference as discussed below.

Ring Current Effects. The term "ring current shifts" is used to denote those chemical shifts in delocalized molecules, relative to an appropriate reference, attributed to the diamagnetic anisotropy resulting from the electron delocalization. **In** discussing ring current shifts it is as important to have a reference chemical shift relative to which the ring current shift is calculated, as to have a diamagnetic reference for calculation of paramagnetic isotropic shifts, discussed above. Unfortunately, satisfactory reference shifts for the species studied in this work are not available. In the following discussion comparison will be made to the free ligand, but it must be kept in mind that bonding the ligand to the metal will change the chemical shifts of the ligand nuclei, and that this shift must be accurately accounted for before comparing the observed shifts to a model for ring current effects.

The chemical shifts of the $2,6$ -C and $3,5$ -C of pyridine in $Ru(CO)(p-i-Pr-TPP)(py)$ are 144.2 and 121.3 ppm, respectively, compared with 149.8 and 123.6 ppm in free pyridine.^{11b} The -5.6 and -2.3 ppm differences between ¹³C resonances in free and coordinated pyridine are similar to the -7.64 and -2.39 ppm shift differences for the 2,6-H and 3,5-H in $Ru(CO)(OEP)(t-Bupy)^{26}$ and -7.09 and -2.08 ppm shift differences for 2,6-H and 3,5-H in $Ru(CO)(p-CF_3-TPP)$ - $(t-Bupp).8$ Changes in shift between free and coordinated pyridine resonances in (py) ₂Co(TPP)Br are -7.80 and -2.08 ppm for 2,6-H and 3,5-H, respectively, and -3.2 and -1.4 ppm for 2,6-C and 3,5-C, respectively.6 13C chemical shifts of pyridine coordinated to a metal have been reported for two platinum complexes: in $(CH_3)_2PtBr_2(py)_2$ the ¹³C shifts for 2,6-C and 3,5-C (CDCl₃ solution, ± 1.5 ppm) are 151.0 and 125.3 ppm, respectively, and in $(CH_3)_2Pt(sal=N-CH_3)(py)$ they are 148.4 and 125.1 ppm, respectively.27 **In** these complexes, which do not have the contribution of porphyrin anisotropy to the chemical shift, the 2,6-C resonance shifts in both directions from the shift in uncoordinated pyridine. These complexes are not presented as reasonable reference materials for calculation of ring current shifts in the metalloporphyrins, but rather as indicators of the uncertainties involved.

The methyl carbon resonance of coordinated $P(OMe)$ ₃ in $Ru(p-CF_3-TPP)(P(OMe))$ ₂ is only -1.2 ppm from the resonance in free P(OMe)3, although in the proton spectrum there is a chemical shift difference of -2.62 ppm between resonances of free and coordinated P(OMe)3. For comparison, note that in $Mo(CO)_{5}(P(OMe)_{3}), W(CO)_{5}(P(OMe)_{3}),$ and Mo- $(CO)_4(P(OMe)_3)_2$ the ¹³C resonance of coordinated $P(OMe)_3$ is shifted 2.0 to 3.1 ppm downfield with respect to the free ligand.28 If a similar downfield component to the chemical shift is induced by coordination of $P(OMe)$ ₃ to the ruthenium porphyrin, the ring current shift is much larger than would be inferred from comparison of the chemical shifts of the methyl carbon in $P(OMe)$ ₃ and $Ru(p-CF_3-TPP)(P(OMe)_{3})_{2}$.

In $Ru(CO)(p\text{-}CF_3\text{-}TPP)(THF)$ the 2,5-C and 3,4-C resonances are -3.7 and -3.3 ppm, respectively, from their positions in free THF.llc The corresponding differences in $Ru(CO)(OEP)(THF)$ are -4.9 and -3.9 ppm. The partial displacement of THF from the coordination sphere of Ru- $(CO)(OEP)(THF)$ upon addition of CO causes a change in chemical shift of $+3.0$ ppm for 3,4-C and $+1.3$ ppm for 2,5-C. The approximate ring current shifts in the ¹H NMR spectra of $Ru(\text{CO})(p\text{-}CF_3\text{-}TPP)$ (THF), using free THF as reference, are -5.4 ppm for 2,5-H and -2.4 ppm for 3,4-H. These results are consistent with previous reports that ring current shifts in 1H and 13C spectra are of similar magnitude.6.29 The greater sensitivity of 13C than IH to changes in bonding, however, make it more difficult to separate ring current effects from other changes that result from coordination to the ruthenium. Thus, bonding influences on the 2,6-C chemical shift in coordinated THF may account for the lower ratio of apparent 2,5-C:3,4-C (1.1) ring current shifts compared to the 2,5-H:3,4-H (2.25) in $Ru(CO)(p$ -CF₃-TPP)(THF).

Even though there are not adequate reference shifts for the determination of ring current shifts for these metalloporphyrin complexes, it is of interest to compare the estimated ring current shifts with calculated values. Among the recent ring current calculations,30-32 direct application to porphyrins is most readily made using the figures in ref 30. Bond distances were obtained from the x-ray crystal structure of Ru(C0)- $(TPP)(py)$ ³³ and $Ru(CO)(TPP)(EtOH)$ ³⁴ or estimated from molecular models. The calculated ring current shifts were extrapolated where necessary, The calculated ring current shifts in ppm and observed shifts relative to the free ligands, in parentheses, for t-Bupy are o-H, 6 (7,l); 0-C, 6.5 *(5.6);* $m-H$, 2 (2.1); $m-C$, 2 (2.3); t-Bu-H, 1 (0.9-1.03), and for P(OMe)3 C, 2 **(3,);** H, 1.7 (2.6). This agreement is probably fortuitous, since it is better than the uncertainty in some of the individual values. Extending this treatment to the carbon of the coordinated CO, the calculations³⁰ extrapolate to an estimated ring current contribution of ca. 10-12 ppm. When this factor is taken into consideration, the 13C shifts of the carbonyl carbon in ruthenium carbonyl porphyrins are consistent with the chemical shifts observed for other Ru-CO units. $21 - 25$

Chemical Shift Differences between Nonequivalent Sites. In indium chloro, titanyl, and ruthenium carbonyl complexes of substituted tetraphenyl porphyrins slow axial ligand exchange and slow rotation, on the NMR time scale, of the phenyl rings with respect to the plane of the porphyrin result in nonequivalence of the two sides of each phenyl ring.8 The resulting chemical shift differences between nonequivalent protons8 are given in Table **111** along with the 13C nonequivalences for the related carbons, in ppm. Chemical shift differences in the proton resonances are for spectra at ca. 30° except for $TiO(p-i-Pr-TPP)$ and $TiO(p-CF_3-TPP)$ where it is necessary to go to 9 and **17O,** respectively, to obtain a slow exchange

Table 111. Chemical Shift Differences (ppm) in ¹H and ¹³C NMR Spectra

A. Nonequivalence Arising from	
Restricted Rotation of Phenyl Rings	

^a The o-H protons are bound to C_2 and C_2' . ^b ¹H spectrum at 17°. ^c ¹H spectrum at 9°. ^d p-H and m-H have similar shifts and individual resonances are not readily assigned.

spectrum.⁸ All ¹³C spectra are at ca. 35°.

For $M(p-CF_3-TPP)X$ the chemical shift differences in ppm between C_2 and C_2 ' in the ¹³C NMR spectra are up to about a factor of **2** larger than the shift differences between their respective protons (0-H and *o-13')* in the IH NMR spectra, Shift differences between C_3 and C_3 ' are not resolvable, due in part to the C-F coupling constant of 3 Hz between $C_3(C_3)$ and the CF3 group. Nonequivalences in the proton spectra are 0.10 ppm or less.⁸ It should also be noted that the ¹H spectra were obtained at 100 MHz and the 13C spectra at 20 MHz and the IH spectra were obtained at higher resolution so that small nonequivalences were much more readily detected in the $\rm{^1H}$ than in the $\rm{^{13}C}$ spectra. The broadening of the C₂ and C_2 ' resonances in the spectrum of TiO(p -CF₃-TPP) is attributed *to* intermediate rate of rotation of the phenyl rings. Considerable broadening of the 1H spectra is also observed at ca. 35°.

The ¹³C spectra of $M(p-i-Pr-TPP)X$ are considerably less informative concerning nonequivalent phenyl sites than are the IH spectra even though the chemical shift differences in the ¹H spectra of M(p-i-Pr-TPP)X and M(p-CF₃-TPP)X are very similar. The observation of a single resonance for C₂ and C_2 ' in TiO(*p-i-Pr-TPP*) is not surprising in view of the ca. 40° coalescence temperature for the ortho protons.8 It is interesting to note that the $13C$ chemical shift differences between C_2 and C_2 ' and between C_3 and C_3 ' in $Ru(CO)(p-i-Pr-TPP)(py)$ are approximately equal, whereas in the 1H NMR of a large range of complexes the nonequivalence between o -H protons was several times larger than between m-H protons.

Striking differences are found between the CH3 resonances of $H_2(o-Me-TPP)$ and $M(o-Me-TPP)X$. In the ¹H NMR spectrum of $H_2(o-Me-TPP)$ the various isomers give rise to five distinguishable CH_3 resonances, but in the ^{13}C spectrum only one CH₃ resonance is seen. The spectra of the $M(o Me$ -TPP)X complexes, however, show ten CH₃ resonances in two major groupings in the **III** spectrum and two CH3 resonances in the 13C spectrum. Similar behavior is observed for the resonances of the ring carbons and their attached protons, except that in this case the 13C spectra are more easily interpreted due to the complicated coupling in the **IH** spectra. These observations imply that the chemical shifts of the o-tolyl ring resonances are dependent primarily upon the orientation of the nucleus relative to the axial ligand on the metal.

Thus there appear to be similarities although not exact correspondance in chemical shift differences (in ppm) between

Thiolato Sulfur as an Electron-Transfer Bridge

nonequivalent sites in the 13C and IH spectra. However, the factor of *5* difference in frequency of observation at comparable field strengths for the two nuclei makes it easier to observe small differences in the **1H** spectra than in the 13C spectra for those cases where the proton-proton coupling pattern is relatively simple.

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Registry No. I, 38350-65-1; **11,** 56398-53-9; **111,** 56398-58-4; IV, 41654-54-0; **V,** 56420-24-7; **VI,** 56398-54-0; VII, 56398-57-3; VIII, 55106-63-3; IX, 57091-08-4; X, 57091-09-5; XI, 37083-40-2; XII, 56398-56-2; XIII, 56398-59-5; XIV, 14074-80-7; XVII, 32125-07-8; XIX, 55059-68-2; Ru(CO)(p-i-Pr-TPP)(EtOH), 55059-74-0; **P-** (OMe)3, 121-45-9; 13C, 14762-74-4.

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Thiolato Sulfur as an Electron-Transfer Bridge. The Chromium(I1)-Catalyzed Aquation of Thiolatobis(ethylenediamine)chromium(III) Complexes in Aqueous Perchloric Acid Media

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The rates of the chromium(II) reductions of the $[Cr(en)(SCH_2COO)]^+$, $[Cr(en)(2SC_6H_4COO)]^+$, $[Cr(en)_2-C_6H_4CO]$ (SCH2CH2NH2)]2+, and [Cr(en)z(OOCCOQ)]+ ions have been measured in aqueous perchloric acid media. All reactions proceed by inner-sphere electron transfer. Reduction of the mercaptoacetato and 2-mercaptobenzoato complexes is strongly catalyzed by acid, whereas reduction of the 2-mercaptoethylamine and oxalato complexes is essentially independent of acid. The acid-catalyzed redox path is interpreted in terms of a proton-induced cis nonbridging ligand effect. Rates of the title reactions are compared to the rates of chromium(I1) reduction of the analogous **thiolatobis(ethylenediamine)cobalt(III)** complexes, and arguments are presented which indicate that the remarkable bridging efficiency reported for thiolato sulfur in **Cr(I1)-Co(I1I)** reactions does not carry **over** to the analogous Cr(II)-Cr(III) reactions. It is suggested that this kinetic difference may arise from the fact that thiolato sulfur induces a ground state trans effect in the cobalt(II1) complexes, but not in an analogous chromium(II1) complex.

Introduction

The remarkable ability of low-valent sulfur to mediate electron transfer from chromium(**11)** to cobalt(111) was first noted by Bennett² for thiolato sulfur. Thiocyanato sulfur was shown to also efficiently bridge electron transfer from chromium(II) to cobalt(III),³ as well as from uranium(III) to chromium(III),4 but a study on electron transfer from cobalt(II) (as $[Co(CN)_5]^{3-}$) to cobalt(III) was inconclusive.⁵ In an effort to determine if the unusually great bridging efficiency of thiolato sulfur in Cr(II)-Co(III) reactions extends to other redox systems, we have prepared, characterized, and quantified the acid-catalyzed aquation of thiolatobis(ethy1 **enediamine)chromium(III)** complexes6 which are analogous to the **thiolatobis(ethylenediamine)cobalt(III)** complexes